ESfEQA
European Society for
External Quality Assessment

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Quality Assessment Schemes Program

2025

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Educational Programs

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ABOUT ESFEQA

A high quality standard is essential for every medical laboratory since test results are the basis for medical decisions and have an important impact on the wellbeing and treatment of patients. There are different approaches to maintain and improve quality in medical laboratories.
ESfEQA – European Society for External Quality Assessment– supports laboratories to assess the quality of their analytical results and ultimately improve their performance by providing well-designed external quality assessment programs.
ESfEQA offers a wide range of External Quality Assessment Schemes.
ESfEQA was founded in Heidelberg/Germany in 2013 and is accredited according to the international standard ISO 17043 by the German national accreditation body DAkkS. Since its foundation, ESfEQA has expanded its program portfolio and at the same time the number of laboratories participating in ESfEQA's external quality assessment schemes.

New Requirements for POCT

With the new version of ISO15189:2022, external quality assurance is also becoming more important for POCT applications. ESfEQA offers the following proficiency tests for POCT applications: Glucose and Prothrombin time (INR), Qualitative urinalysis (urine strips), hCG in urine and serum, Covid antigen tests, Erythrocyte Sedimentation Rate, Blood Gas & Electrolytes, CO-Oximetry, Drugs in Urine, Fecal Occult Blood (FOB) and Dengue N1 Antigen.

We look forward to your feedback and ideas for further EQA schemes in the POCT area.

Registration and Sample Ordering

ESfEQA offers EQA schemes worldwide and cooperates with reliable regional distributors. They are the direct business partners of participants, responsible for the ordering process, invoicing and local shipment of survey samples.
ESfEQA offers programs with 2, 4 or 12 surveys per year. In general, programs should be ordered for an entire calendar year.

Survey Calendar

The dates for begin of result entry and deadline for result entry are published in this catalogue and on the ESfEQA website (www.esfeqa.eu).
The testing periods of the proficiency test programs are synchronized in order to make the samples of a year available to participants in as few shipments as possible. Thus, a maximum of 4 shipments per year are required per participant.
Survey samples are sent to the participants in good time, usually at the beginning of the respective testing period. In order to keep the logistical, environmental and financial effort as low as possible, the samples of the first and second quarters of the monthly and quarterly programs are sent together; as well as the samples for the third and fourth quarters. This procedure is chosen for samples with sufficient stability for a period of at least 6 months. Samples with a shorter shelf life, as well as the samples for the semi-annual programs are sent quarterly.

Submission of Results, Survey Reports and Certificates

Participants submit their results online via the TEQA web-application. Requests for new method, instrument and reagent codes can be made online. The subscription to any ESfEQA program allows participants to submit up to three results obtained from a single control set using different devices. Reports and certificates are provided online as pdf-files within 3 weeks (within 10 days for monthly programs) after the deadline of result submission. Report files and certificates can be stored electronically, forwarded and printed.

New Programs

Based on the feedback of our participants, ESfEQA extends the EQA portfolio continously. Please contact us for further suggestions on new programs.
Programs that have not been listed on the ESfEQA ISO 17043 accreditation certificate yet are marked in this catalogue as not accredited.

Heidelberg, August 2024

BIOCHEMISTRY PROGRAMS

BILIRUBIN NEONATAL — BILI-N

Program: BILI-N: 4 surveys/year x 2 samples
Material: Lyophilized samples of human Serum (minimum 0,5mL)
Evaluation: Quantitative

Analytical parameters:

Bilirubin direct	Bilirubin total
Bilirubin conjugated	Bilirubin non-conjugated

BLOOD GAS AND ELECTROLYTES — BG

New: Magnesium

Program: BG12: 12 surveys/year x 1 sample
BG4: 4 surveys/year x 2 samples
Material: Liquid buffered aqueous solution or serum-based samples (minimum 2 mL)
Evaluation: Quantitative

Analytical parameters:

Bicarbonate (HCO_3^-)	Lactate	pO2
Calcium	Magnesium	Potassium
Chloride	pCO2	Sodium
Glucose	pH	Urea

CARDIAC MARKER — CM

Program: CM12: 12 surveys/year x 1 sample
CM4: 4 surveys/year x 2 samples
CM2: 2 surveys/year x 2 samples
Material: Lyophilized samples of human sera with added analytes of human origin (minimum 1 mL)
Evaluation: Quantitative

Analytical devices that are intended for whole blood only are not suitable for these samples.

Analytical parameters:

BNP	Homocysteine	Troponin I
CK-MB (mass)	Myoglobin	Troponin T
CK-MB (activity)	NT-proBNP	

CLINICAL CHEMISTRY CC

Program: CC12: 12 surveys/year x 1 sample
CC4: 4 surveys/year x 2 samples
CC2: 2 surveys/year x 2 samples

Material: Lyophilized samples of human sera with added enzymes and proteins of human origin (5 mL)
Evaluation: Quantitative

Analytical parameters:

Albumin	Cholesterol	Lithium
ALP Alkaline phosphatase	Cholinesterase	Magnesium
ALT/GPT	CK Creatinkinase	Phosphate
α-Amylase	Creatinine	Potassium
Amylase pancreatic	Copper	Sodium
AST/GOT	Gamma GT	TIBC Total Iron Binding Capacity
Bilirubin, direct	Glucose	Total protein
Bilirubin, total	HDL Cholesterol	Triglycerides
Bilirubin conjugated	Iron	UIBC Unsaturated Iron Binding Capacity
Bilirubin non-conjugated	Lactate	Urea
Calcium	LDH Lactate Dehydrogenase	Uric acid
Calcium (ionized)	LDL Cholesterol	Zinc
Chloride	Lipase	

COAGULATION COA

Program: COA12: 12 surveys/year x 1 sample
COA4: 4 surveys/year x 2 samples
COA2: 2 surveys/year x 2 samples

Material: Lyophilized samples of human plasma (1 mL)
Evaluation: Quantitative

Analytical parameters:

aPTT (activated Partial Thromboplastin Time)	D-Dimer	Protein C
Antithrombin III	Fibrinogen	Protein S
	PT (prothrombin time)	Thrombin Time

CO-OXIMETRY OXI

Program: OXI4: 4 surveys/year x 2 samples

Material: Purified bovine hemoglobin solution treated with carbon monoxide (minimum 1,0 mL)
Evaluation: Quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043

Analytical parameters:

Oxyhemoglobin	Carboxyhemoglobin	total Hemoglobin
Desoxyhemoglobin	Methemoglobin	

CSF DIAGNOSTICS — CSF

Program: CSF4: 4 surveys/year x 2 samples

Material: Liquid samples made from human serum and other human and chemical components (minimum 1 mL)
Evaluation: Quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043

Analytical parameters:

Albumin	IgG	
Chloride	IgM	Sodium
Glucose	Lactate	Proteine
IgA	LDH	

DRUGS OF ABUSE — DAT

New: Fentanyl Phencyclidine

Program: DAT: 4 surveys/year x 2 samples

Material: Liquid or lyophilized samples of filtered human urines with added drugs for qualitative analysis (minimum 1 mL)
Evaluation: Qualitative

Analytical parameters:

Acetylmorphine	Cannabinoids	Metamphetamines
Amphetamines	Cocaine and metabolites	Opiates
Barbiturates	Fentanyl*	Phencyclidine*
Benzodiazepines	MDMA	Synthetic Cannabinoids (K2/Spice)
Buprenorphine	Methadone and metabolites	Tricyclic Antidepressants

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

ETHANOL, AMMONIA AND BICARBONATE — ETH

Program: ETH12: 12 surveys/year x 1 sample
ETH4: 4 surveys/year x 2 samples

Material: Liquid samples with added compounds (minimum 1 mL)
Evaluation: Quantitative

Analytical parameters:

Ethanol	Ammonia	Bicarbonate

FECAL OCCULT BLOOD — FOB

Program: FOB: 2 surveys/year x 2 samples

Material: Liquid samples simulating extracted stool samples (minimum 0,5 mL)
Evaluation: Qualitative and quantitative

Analytical parameters:

Human Hemoglobin (qualitative and quantitative)

GLUCOSE POC - WHOLE BLOOD — GLUWB

Program: 4 surveys/year x 2 samples
GLUWB: Registration of 1-3 measuring systems
GLUWB 6 DEVICES: Registration of up to 6 measuring systems
GLUWB 9 DEVICES: Registration of up to 9 measuring systems

Material: Simulated whole blood (minimum 1 mL)
Evaluation: Quantitative

Analytical parameters:

Glucose

GLYCATED HEMOGLOBIN (HbA1c) — GHB

Program: GHB12: 12 surveys/year x 1 sample
GHB4: 4 surveys/year x 2 samples

Material: Lyophilized samples of hemolysate of human blood (minimum 0,5 mL)
Evaluation: Quantitative

Analytical parameters:

HbA1c

PROTHROMBIN TIME (INR)-POCT — INR-POCT

Program: 4 surveys/year x 2 samples
INR-POCT: Registration of 1-3 measuring systems
INR-POCT 6 DEVICES: Registration of up to 6 measuring systems
INR-POCT 9 DEVICES: Registration of up to 9 measuring systems

Material: Liquid samples (minimum 0,3 mL)
Evaluation: Quantitative

Suitable for POCT analyzers, e.g. Roche Coaguchek, Siemens Xprecia Stride, Abbott iStat.

Analytical parameters:

Prothrombin Time (INR)

QUALITATIVE URINE ANALYSIS (URINE STICK) — US

Program: US4: 4 surveys/year x 2 samples
US2: 2 surveys/year x 2 samples

Material: Liquid samples of urine preparation of human origin with added preservatives and stabilizers (minimum 10 mL)
Evaluation: Semi-quantitative

Analytical parameters:

Bilirubin	Ketone bodies	Specific Gravity
Glucose	Leucocytes	Total Protein
hCG	Nitrite	Urobilinogen
Hemoglobin	pH	

THERAPEUTIC DRUGS — TDM

Program: TDM**:** 4 surveys/year x 2 samples

Material: Liquid samples with added compounds (minimum 2 mL)
Evaluation: Quantitative

Analytische Parameter:

Amikacin	Gentamicin	Procainamide
Carbamazepine	Lidocain	Salicylate
Chinidine	NAPA	Theophylline
Chloramphenicol	Paracetamol	Tobramycin
Digoxin	Phenobarbital	Valproic Acid
Disopyramide	Phenytoin	Vancomycin
Ethosuximide	Primidone	

URINE CHEMISTRY — UC

Program: UC: 4 surveys/year x 2 samples

Material: Liquid or lyophilized samples of human urine (minimum 5 mL)
Evaluation: Quantitative

Analytical parameters:

Albumin / Microalbumin	Glucose	Total protein
Amylase*	Magnesium	Sodium
Calcium	Osmolality*	Urea
Chloride	Phosphate	Uric acid
Creatinine	Potassium	

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

URINE SEDIMENT FOR LIGHT SCATTERING METHODS — USEDL

Program: USEDL4: 4 surveys/year x 2 samples
USEDL2: 2 surveys/year x 2 samples

Material: Liquid samples of human urine (minimum 5 mL)
Evaluation: Qualitative, quantitative and semi-quantitative

This program is suitable for light scattering methods, e.g. Sysmex UF-5000/4000/1500.

Analytical parameters:

Bacteria qual., semi-quant., quant.	Red cells qual., semi-quant., quant.
Casts qual., semi-quant., quant.	White cells qual., semi-quant., quant.
Crystals qual., semi-quant., quant.	

URINE SEDIMENT FOR MICROSCOPIC METHODS USEDM

Program: USEDM4: 4 surveys/year x 2 samples
USEDM2: 2 surveys/year x 2 samples

Material: Liquid samples of human urine (minimum 5 mL)
Evaluation: Qualitative, quantitative and semi-quantitative

This program is suitable for manual microscopy and automated microscopy, e.g. Siemens Atellica, Beckman Coulter Iris, Roche Cobas u 701, Menarini Sedimax, 77 Elektronika UriSed, Dirui FUS, Analyticon Urilyzer Cell, Mindray EH Series, Mindray EU series.

Analytical parameters:

Bacteria qual., semi-quant., quant.
Casts qual., semi-quant., quant.
Crystals qual., semi-quant., quant.
Red cells qual., semi-quant., quant.
White cells qual., semi-quant., quant.

IMMUNOLOGY PROGRAMS

HCG IN SERUM — HCG

Program: HCG: 4 surveys/year x 1 sample

Material: Lyophilized or liquid sample of human serum with added analytes of human origin (minimum 1 mL)
Evaluation: Qualitative

Analytical parameters:

hCG qualitativ

HCG IN URINE — HCGU

Program: HCGU: 4 surveys/year x 2 samples

Material: Liquid samples of synthetic urine (minimum 1 mL)
Evaluation: Qualitative

Analytical parameters:

hCG qualitativ

HORMONES — HOR

Program: HOR12: 12 surveys/year x 1 sample
HOR4: 4 surveys/year x 2 samples

Material: Lyophilized samples of human sera with added analytes (minimum 3 mL)
Evaluation: Quantitative

Analytical parameters:

Aldosterone	hCG	SHBG
AMH	Homocysteine	T3, free
Androstenedione	Human Growth Hormone	T3, total
Calcitonin	IgE	T4, free
C-Peptide	IGF-1*	T4, total
Cortisol	Insulin	Testosterone
DHEA-S	LH (Luteinizing Hormone)	Thyroglobulin
Estradiol	Methylmalonic Acid	TSH
Ferritin	PTH	Vitamin B12
Folate	Progesterone	Vitamin D (25-OH)
FSH	Prolactin	17-OH-Progesterone

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

PROCALCITONIN — PCT

Program: PCT: 4 surveys/year x 2 samples

Material: Lyophilized samples of human sera with added analyte (minimum 0,5 mL)
Evaluation: Quantitative

Analytical parameters:

Procalcitonin

SPECIFIC PROTEINS — SP

Program: SP12: 12 surveys/year x 1 sample
SP4: 4 surveys/year x 2 samples

Material: Liquid or lyophilized samples of human sera with added analytes of human origin (minimum 1 mL)
Evaluation: Quantitative

Analytical parameters:

Albumin	Ceruloplasmin	Prealbumin
Alpha-1-acid glycoprotein	CRP (C-Reactive Protein)	RF
Alpha-1-antitrypsin	Cystatin C*	soluble Transferrin receptor
Alpha-2-macroglobulin	Haptoglobin	(sTfR)*
ASO	IgA, IgE, IgG, IgM	Transferrin
Beta-2-microglobulin	Kappa light chains, total* and free	
C3, C4	Lambda light chains, total* and free	

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

THYROID ANTIBODIES — ANTI-THYR

Program: ANTI-THYR: 4 surveys/year x 2 samples

Material: Samples liquid or lyophilized (0,5 mL)
Evaluation: Quantitative

Analytical parameters:

anti-TG	anti-TPO
TRAb (TSH-Receptor Antibodies)	

TUMOR MARKER — TM

Program: TM12: 12 surveys/year x 1 sample
TM4: 4 surveys/year x 2 samples

Material: Lyophilized samples of human sera with added analytes (minimum 3 mL)
Evaluation: Quantitative

Analytical parameters:

AFP	CA 125	PSA, total
CEA	CA 15-3	PSA, free
CA 19-9	Ferritin	

TUMOR MARKER & HORMONES TMH

Program: TMH12: 12 surveys/year x 1 sample
TMH4: 4 surveys/year x 2 samples
TMH2: 2 surveys/year x 2 samples

Material: Lyophilized samples of human sera with added analytes (minimum 3 mL)
Evaluation: Quantitative

Analytical parameters:

AFP	Folate	PTH
Aldosterone	FSH	SHBG
AMH	hCG	T3, free
Androstenedione	Homocysteine	T3, total
CA 125	Human Growth Hormone	T4, free
CA 15-3	IgE	T4, total
CA 19-9	IGF-1*	Testosterone
Calcitonin	Insulin	Thyroglobulin
CEA	LH (Luteinizing Hormone)	TSH
Cortisol	Methylmalonic Acid	Vitamin B12
C-Peptide	Progesterone	Vitamin D (25-OH)
DHEA-S	Prolactin	17-OH-Progesterone
Estradiol	PSA, free	
Ferritin	PSA, total	

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

MICROBIOLOGY PROGRAMS

ADENOVIRUS SEROLOGY — ADE

Program: ADE: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

The scheme is intended for Novatec, Virion/Serion, Virotech and Euroimmun ELISA reagents. Other reagents upon request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgA, IgG and IgM antibodies against Adenovirus

ASPERGILLUS FUMIGATUS SEROLOGY — ASF

Program: ASF: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

The scheme is intended for Virion/Serion ELISA reagents. Other reagents upon request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgA, IgG, IgM and total antibodies against Aspergillus fumigatus

ASPERGILLUS GALACTOMANNAN ANTIGEN — ASPAG

Program: ASPAG: 2 surveys/year x 2 samples

Material: Liquid samples of simulated bronchoalveolar lavage (BAL) fluid or serum (minimum 0,5 mL)
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Aspergillus Antigen (Galactomannan)

BACTERIOLOGY — BAC-C, BAC-E

Program: BAC-C or BAC-E: 4 surveys/year x 4 samples

Material: Lyophilised samples (pure strain and/or mixture of bacteria): 2 for identification and 2 for antibiotic susceptibility testing (AST). AST according to EUCAST or CLSI guidelines.

In this program we simulate different types of specimens: blood, urine, swabs (e.g. surgical/wound site, etc.), sputum/bronchospcopy specimen, paracentesis samples (e.g. ascites), joint/synovial fluid, sonicate fluid of explanted prosthetic joints, and CSF.
Evaluation: Qualitative

Analytical parameters:

Identification (genus and species)
Antibiotic susceptibility testing (according to EUCAST or CLSI guidelines)

BORDETELLA SEROLOGY — BPES

Program: BPES: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Bordetella IgA, IgG, IgM	Bordetella Pertussis-Toxin IgA	Bordetella Pertussis-Toxin IgG

BORRELIA SEROLOGY — BOR

Program: BOR: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgG and IgM antibodies against Borrelia burgdorferi

BORRELIA IgG ANTIBODY INDEX — BOR-G-AI

Program: BOR-G-AI: 2 surveys/year x 2 samples
Material: One CSF/serum sample pair and (simulated) clinical information on the patient needed to calculate the antibody index is provided to the participant (CSF sample: 0,8 mL; serum sample: 0,3 mL)
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Borrelia IgG-antibody index (AI), qualitative and quantitative

BORRELIA IgM ANTIBODY INDEX — BOR-M-AI

Program: BOR-M-AI: 2 surveys/year x 2 samples
Material: One CSF/serum sample pair and (simulated) clinical information on the patient needed to calculate the antibody index is provided to the participant (CSF sample: 0,8 mL; serum sample: 0,3 mL)
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Borrelia IgM-antibody index (AI), qualitative and quantitative

BRUCELLA SEROLOGY — BRU

Program: BRU: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgA, IgG and IgM antibodies against Brucella | agglutinating antibodies against Brucella

CHAGAS SEROLOGY — CHA

Program: CHA: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgG and IgM antibodies against Trypanosoma cruzi

CHIKUNGUNYA VIRUS SEROLOGY — CHIKV

Program: CHIKV: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgG and IgM antibodies against Chikungunya Virus

CHLAMYDIA TRACHOMATIS SEROLOGY — CHT

Program: CHT: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgA, IgG and IgM antibodies against Chlamydia trachomatis

COXSACKIEVIRUS SEROLOGY — COX

Program: COX: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

The scheme is intended for Virion/Serion ELISA and Euroimmun IFT reagents. Other reagents upon request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgA, IgG and IgM antibodies against Coxsackievirus

DENGUE VIRUS ANTIBODIES — DENV

Program: DENV: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgG and IgM antibodies against Dengue Virus

DENGUE VIRUS NS1 ANTIGEN — DENVAG

Program: DENVAG: 2 surveys/year x 2 samples

Material: Liquid or lyophilized samples. The samples are either serum or plasma samples or simulated samples consisting of an aqueous protein matrix. Dengue virus NS1 antigen positive samples contain recombinant DENV NS1 protein
Evaluation: Qualitative

This programme is intended for immunochromatographic tests (Lateral Flow Rapid tests) and ELISA. Other reagents on request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Dengue Virus NS1 antigen

ECHO VIRUS SEROLOGY — ECH

Program: ECH: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

The scheme is intended for Virion/Serion ELISA and Euroimmun IFT reagents. Other reagents upon request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgA, IgG and IgM antibodies against ECHO-Virus

ENTEROVIRUS SEROLOGY — ENT

Program: ENT: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

The scheme is intended for Virion/Serion, Virotech and Euroimmun ELISA reagents. Other reagents upon request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgA, IgG and IgM antibodies against Enterovirus

EPSTEIN-BARR VIRUS SEROLOGY — EBV

Program: EBV: 4 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

anti-EBV EBNA-1 IgG + total	anti-EBV VCA IgG + total	anti-EBV VCA IgM

HELICOBACTER PYLORI ANTIBODIES — HPYL

Program: HPYL: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Helicobacter pylori IgA, IgG, IgM and total antibodies

HEPATITIS A VIRUS SEROLOGY — HAV

Program: HAV: 4 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

anti-HAV IgG + total	anti-HAV IgM

HEPATITIS B VIRUS SEROLOGY — HBV

Program: HBV: 4 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 1 mL)
Evaluation: Qualitative and quantitative

Analytical parameters:

anti-HBs (qual. and quant.*)	anti-HBe	HBsAg (qual. and quant.*)
anti-HBc IgG + total	anti-HBc IgM	HBeAg

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

HEPATITIS E VIRUS SEROLOGY — HEV

Program: HEV: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

anti-HEV IgG + total	anti-HEV IgM

HIV ANTIBODIES AND ANTIGEN — HIV

Program: HIV: 4 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative
Please note that the HIV survey is designed for assays that detect HIV antibodies and HIV antigen separately. For combo tests (e.g. HIV 4th generation assays) that detect HIV antibodies and HIV antigen simultaneously we recommend the enrollment in the ESfEQA INF survey.

Analytical parameters:

anti-HIV 1/2 antibodies	HIV p24 Antigen*

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

HTLV I/II — HTL

Program: HTL: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

anti-HTLV I/II

INFECTIOUS DISEASE COMBINATION CONTROL SEROLOGY — INF

Program: INF4: 4 surveys/year x 2 samples
INF4x4: 4 surveys/year x 4 samples
INF2: 2 surveys/year x 2 samples
INF12: 12 surveys/year x 1 sample

New Program INF12

Material: Liquid samples of defibrinated human plasma (minimum 1 mL)
Evaluation: Qualitative

Analytical parameters:

anti-HIV 1/2 / p24 Ag	anti-HBc
anti-HCV	HBsAg

INFLUENZA A VIRUS SEROLOGY — INA

Program: INA: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

The scheme is intended for Novatec, Virion/Serion, Virotech and Euroimmun ELISA reagents. Other reagents upon request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgA, IgG and IgM antibodies against Influenza A Virus

INFLUENZA B VIRUS SEROLOGY — INB

Program: INB: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

The scheme is intended for Novatec, Virion/Serion, Virotech and Euroimmun ELISA reagents. Other reagents upon request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgA, IgG and IgM antibodies against Influenza B Virus

INFLUENZA A ANTIGEN — FLUAAG

Program: FLUAAg: 2 surveys/year x 2 samples

Material: Liquid or lyophilized samples simulating swab specimens (e.g. oropharyngeal, nasopharyngeal, nasal etc.). Influenza A antigen positive samples contain inactivated whole virus (minimum 0,3 mL).
Evaluation: Qualitative

New Program

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Influenza A Antigen

INFLUENZA B ANTIGEN — FLUBAG

Program: FLUBAg: 2 surveys/year x 2 samples

Material: Liquid or lyophilized samples simulating swab specimens (e.g. oropharyngeal, nasopharyngeal, nasal etc.). Influenza B antigen positive samples contain inactivated whole virus (minimum 0,3 mL).
Evaluation: Qualitative

New Program

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Influenza B Antigen

LEGIONELLA PNEUMOPHILA ANTIBODIES — LPAB

Program: LPAB: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgG, IgM and total antibodies against Legionella pneumophila

LEPTOSPIRA SEROLOGY LEP

Program: LEP: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgG and IgM antibodies against Leptospira	agglutinating antibodies against Leptospira*

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

MALARIA MICROSCOPY MALM

Program: MALM: 4 surveys/year x 2 samples
Material: Slides of stained smears
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Malaria Parasite Detection	Stage Identification
Species Identification	Quantification of Plasmodium falciparum

MEASLES SEROLOGY MEA

Program: MEA: 2 surveys/year x 2 samples
Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgG and IgM antibodies against Measles Virus

MYCOPLASMA ANTIBODIES MYPL

Program: MYPL: 2 surveys/year x 2 samples
Material: Samples of human serum (minimum 0,3 mL)
Evaluation: Qualitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Mycoplasma pneumoniae IgA, IgG, IgM and total antibodies

PARAINFLUENZA VIRUS SEROLOGY — PIN

Program: PIN: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

The scheme is intended for Novatec, Virion/Serion, Virotech and Euroimmun ELISA reagents. Other reagents upon request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgA, IgG and IgM antibodies against Parainfluenza Virus

PARVOVIRUS B19 SEROLOGY — PAR

Program: PAR: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgG and IgM antibodies against Parvovirus B19

RESPIRATORY SYNCYTIAL VIRUS (RSV) ANTIGEN — RSVAG

Program: RSVAg: 2 surveys/year x 2 samples

New Program

Material: Liquid or lyophilized samples simulating swab specimens (e.g. oropharyngeal, nasopharyngeal, nasal etc.). RSV antigen positive samples contain inactivated whole virus (minimum 0,3 mL).
Evaluation: Qualitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Respiratory Syncytial Virus Antigen

RESPIRATORY SYNCYTIAL VIRUS (RSV) SEROLOGY — RSV

Program: RSV: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

The scheme is intended for Novatec, Virion/Serion, Virotech and Euroimmun IFT reagents. Other reagents upon request.

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

IgG, IgM and IgA antibodies against Respiratory Syncytial Virus (RSV)

RESPIRATORY VIRAL ANTIGEN DETECTION — RESPAG

New Program

Program: RESPAg: 2 surveys/year x 3 samples

Material: Lyophilized samples (minimum 0,3mL) simulating swab specimens (e.g. oropharyngeal, nasopharyngeal, nasal etc.) or swabs. Antigen positive samples contain inactivated whole virus.
Evaluation: Qualitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Flu A Antigen	Flu B Antigen	RSV Antigen

SARS-CoV-2 ANTIGEN — COVAG

Program: COVAG: 4 surveys/year x 3 samples

Material: Liquid or lyophilized samples simulating swab specimens (e.g. oropharyngeal, nasopharyngeal, nasal etc.). SARS-CoV-2 antigen positive samples contain inactivated whole virus (minimum 0,3 mL).
Evaluation: Qualitative

Analytical parameters:

SARS-CoV-2 Antigen (qualitative)

SARS-CoV-2 SEROLOGY — COVID

Program: COVID: 2 Surveys/year x 4 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

Analytical parameters:

IgA, IgG, IgM and total antibodies against SARS-CoV-2
SARS-CoV-2 neutralising antibodies

STREPTOCOCCUS A ANTIGEN — STAA

Program: STAA: 2 Surveys/year x 2 samples

Material: Swab
Evaluation: Qualitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Streptococcus A Antigen

SYPHILIS SEROLOGY — SYP

Program: SYP4: 4 surveys/year x 2 samples
SYP2: 2 surveys/year x 2 samples
SYP12: 12 surveys/year x 1 sample

New Program SYP12

Material: Liquid samples of defibrinated human plasma (1 mL)
Evaluation: Qualitative and quantitative

Analytical parameters:

anti-Treponema pallidum antibodies (qualitative)
anti-Treponema pallidum antibodies (semiqualitative)
IgG and IgM antibodies against Treponema pallidum (qualitative)*
Non-treponemal Lipoid antibodies (RPR/VDRL Tests) (qualitative)
Non-treponemal Lipoid antibodies (RPR/VDRL Tests Titers) (semi-quantitative)*

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

TBEV IgG ANTIBODY INDEX — TBEV-G-AI

Program: TBEV-G-AI: 2 surveys/year x 2 samples

Material: CSF/serum sample pair and (simulated) clinical information on the patient needed to calculate the antibody index is provided to the participant (CSF sample 0,8 mL; 0,3 mL for the serum sample)
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

TBEV IgG-antibody index (AI), qualitative and quantitative

TBEV IgM ANTIBODY INDEX — TBEV-M-AI

Program: TBEV-M-AI: 2 surveys/year x 2 samples

Material: CSF/serum sample pair and (simulated) clinical information on the patient needed to calculate the antibody index is provided to the participant (CSF sample 0,8 mL; 0,3 mL for the serum sample)
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

TBEV IgM-antibody index (AI), qualitative and quantitative

ToRCH SEROLOGY — TORCH

Program: TORCH: 4 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 1 mL)
Evaluation: Qualitative and quantitative

Analytical parameters:

anti-CMV IgG (qual. and quant.*)
anti-CMV IgM
anti-HSV 1/2 IgG (qual. and quant.*)
anti-HSV 1/2 IgM
anti-HSV 1 IgG
anti-HSV 2 IgG
anti-HSV 1 IgM
anti-HSV 2 IgM
anti-Rubella IgG (qual. and quant.*)
anti-Rubella IgM
anti-Toxoplasmose gondii IgG (qual. and quant.*)
anti-Toxoplasma gondii IgM

* The quantitative antibody determination is not accredited according to DIN EN ISO/ IEC 17043.

VARIZELLA ZOSTER VIRUS SEROLOGY — VZV

Program: VZV: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative and quantitative

Analytical parameters:

IgA, IgG, and IgM antibodies against Varizella Zoster Virus (VZV), qual. and quant.*

* The quantitative antibody determination is not accredited according to DIN EN ISO/ IEC 17043.

WEST NILE VIRUS SEROLOGY — WNV

Program: WNV: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgG and IgM antibodies against West Nile Virus

ZIKA VIRUS SEROLOGY — ZIKV

Program: ZIKV: 2 surveys/year x 2 samples

Material: Liquid samples of defibrinated human plasma (minimum 0,3 mL)
Evaluation: Qualitative

Analytical parameters:

IgG and IgM antibodies against Zika Virus

HBV MOLECULAR — HBVM

Program: HBVM: 4 surveys/year x 3 samples

Material: Lyophilized samples of human serum. Virus positive samples contain the whole genome of inactivated HBV (minimum 1 mL)
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

HBV-DNA (qualitative and quantitative)

HCV MOLECULAR — HCVM

Program: HCVM: 4 surveys/year x 3 samples

Material: Lyophilized samples of human serum. Virus positive samples contain the whole genome of inactivated HCV (minimum 1 mL)
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

HCV-RNA (qualitative and quantitative)

HIV MOLECULAR — HIVM

Program: HIVM: 4 surveys/year x 3 samples

Material: Lyophilized samples of human serum. Virus positive samples contain the whole genome of inactivated HIV (minimum 1 mL)
Evaluation: Qualitative and quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

HIV-RNA (qualitative and quantitative)

SARS-COV-2 MOLECULAR — COVM

Program: COVM: 4 surveys/year x 3 samples

Material: Liquid or lyophilized samples containing human epithel cells or fibroblasts as control for positive nucleic acid extraction and amplification. Virus-positive samples contain the whole genome of inactivated SARS-CoV-2, thus covering all possible gene targets used in different NAT/PCR assays (minimum 1 mL).
Evaluation: Quantitative

Analytical parameters:

SARS-CoV-2 RNA (qualitative) General detection as well as reporting per gene target	SARS-CoV-2 RNA (quantitative) General indication as well as reporting of quantitative value per gene target

HEMATOLOGY PROGRAMS

BLOOD GROUPING — ABO

Program: AB0: 4 surveys/year x 2 samples

Material: Liquid samples of stabilized human red cells suspended in a buffered fluid and preservative. Erythrocyte suspensions contain a red blood cell concentration of 8% minimum (minimum 4 mL).
Evaluation: Qualitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

ABO-Typing	Rhesus (D)-Detection

ERYTHROCYTE SEDIMENTATION RATE — ESR

Program: ESR: 4 surveys/year x 2 samples

Material: Liquid samples containing erythrocytes in blood collection tubes (75x13mm) with pierceable caps (3 mL)
Evaluation: Quantitative

The samples are not suitable for testing on Alifax and Alcor iSED instruments.

Analytical parameters:

Erythrocyte Sedimentation Rate

ERYTHROCYTE SEDIMENTATION RATE FOR ALCOR — ESRAL

Program: ESRAL: 2 surveys/year x 2 samples

Material: Liquid samples of stabilized human red cells suspended in a buffered fluid and preservative (4 mL)
Evaluation: Quantitative

Analytical parameters:

Erythrocyte Sedimentation Rate

ERYTHROCYTE SEDIMENTATION RATE FOR ALIFAX — ESRAF

Program: ESRAF-G: 2 surveys/year x 3 samples in Greiner tubes
ESRAF-S: 2 surveys/year x 3 samples in Sarstedt tubes

Material: Liquid samples for transmittance measurement related to ESR values in human samples (3 mL)
Evaluation: Quantitative

This program is not accredited according to DIN EN ISO/ IEC 17043.

Analytical parameters:

Erythrocyte Sedimentation Rate

HEMOGRAM HEM

Program: HEM12: 12 surveys/year x 1 sample
HEM4: 4 surveys/year x 2 samples
HEM2: 2 surveys/year x 2 samples

Material: Plasma like fluid samples that contain stabilized human red blood cells, white blood cells and platelets of human and/or non-human analogs (minimum 2 mL)
Evaluation: Quantitative

Analytical parameters:

HCT (hematocrit)
HGB (hemoglobin)
MCH (mean corpuscular hemoglobin)
MCHC (mean cellular hemoglobin concentration)
MCV (mean corpuscular volume)
MPV (mean platelet volume)
PCT (Plateletcrit)
PLT (platelets)
RBC (red blood cells)
RDW (RBC distribution width)
WBC (white blood cells)

HEMOGRAM INCL. 3-PART DIFF. HEM3D

Program: HEM3D: 4 surveys/year x 2 samples

Material: Plasma like fluid samples that contain stabilized human red blood cells, white blood cells and platelets of human and/or non-human analogs (minimum 1,5 mL)
Evaluation: Qualitative

This program is dedicated for 3-part WBC/leucocyte differential hematology analyses

Analytical parameters:

GRAN (granulocytes)
HCT (hematocrit)
HGB (hemoglobin)
LYMPH (lymphocytes)
MCH (mean corpuscular hemoglobin)
MCHC (mean cellular hemoglobin concentration)
MCV (mean corpuscular volume)
MID, MXD (mid-sized leucocytes)
MONO (monocytes)
MPV (mean platelet volume)
NEUT (Neutrophiles)
PCT (plateletcrit)
PLT (platelets)
RBC (red blood cells)
RDW (RBC distribution width)
WBC (white blood cells)

HEMOGRAM INCL. 5-PART DIFF. HEM5D

Program: HEM5D12: 12 surveys/year x 1 sample
HEM5D4: 4 surveys/year x 2 samples

Material: Plasma like fluid samples that contain stabilized human red blood cells, white blood cells and platelets of human and/or non-human analogs (minimum 1,5 mL)
Evaluation: Quantitative

This program is dedicated for 5-part WBC/leucocyte differential hematology analyses

Analytical parameters:

BASO (basophiles)*
EO (eosinophiles)*
HCT (hematocrit)
HGB (hemoglobin)
LYMPH (lymphocytes)
MCH (mean corpuscular hemoglobin)
MCHC (mean cellular hemoglobin concentration)
MCV (mean corpuscular volume)
MONO (monocytes)
MPV (mean platelet volume)
NEUT (neutrophiles)
PCT (plateletcrit)
PDW (platelet distribution width)*
PLT (platelets)
RBC (red blood cells)
RDW (RBC distribution width)
RET (reticulocytes)
WBC (white blood cells)
IG (Immature Granulocytes)*

* This parameter is not accredited according to DIN EN ISO/ IEC 17043.

IMMUNOHEMATOLOGY — IMHEM

Program: IMHEM: 2 surveys/year x 6 samples

Material: 2 Erythrocyte suspension (patient; min. 4 mL), 2 serum sample (patient; min. 4 mL) and 2 erythrocyte suspensions (donor; min. 4 mL). Erythrocyte suspensions contain a red blood cell concentration of 8% minimum

Evaluation: Qualitative

Analytical parameters:

ABO-Typing	Rh-Typing	Antibody screening
A-Subtypes	Kell-Antigen Detection	Antibody identification
Rhesus (D)-Detection	Direct Coombs test	Cross-matching

EDUCATIONAL PROGRAMS

CLINICAL CASE STUDY PROGRAM — CASE

For new Subscribers: 2 cases free of charge

12 cases/year

This program focuses on the interpretation of analytical data and aims to support and strengthen the skills of staff to draw the right conclusions from the analytical results.
Participants receive the case description online and submit their interpretation of the clinical data via the ESfEQA web application.

Parameter:

Suspected diagnosis	Parameters supporting the suspected diagnosis
Other tests to confirm the diagnosis	Therapy suggestions

CASE STUDIES IN CLINICAL LABORATORY SCIENCE — CASE-T

For new Subscribers: 2 cases free of charge

6 cases/year

The target group of this program is technical personnel as well as laboratory doctors in medical laboratories. It aims to support and to strengthen the skills of the staff for (pre)analytical questions.
Participants receive the case description online and submit their interpretation of the clinical data via the ESfEQA web application.

Monthly	Program / Date*
	04/02/2025 - 18/02/2025
	25/02/2025 - 11/03/2025
	18/03/2025 - 01/04/2025
	22/04/2025 - 06/05/2025
	20/05/2025 - 03/06/2025
	17/06/2025 - 01/07/2025
	22/07/2025 - 05/08/2025
	19/08/2025 - 02/09/2025
	16/09/2025 - 30/09/2025
	21/10/2025 - 04/11/2025
	11/11/2025 - 25/11/2025
	02/12/2025 - 16/12/2025
BG12	Blood Gas and Electrolytes
CASE	Clinical Case Study Program
CC12	Clinical Chemistry
CM12	Cardiac Marker
COA12	Coagulation
ETH12	Ethanol, Ammonia and Bicarbonate
GHB12	Glycated Hemoglobin (HbA1c)
HEM12	Hemogram
HEM5D12	Hemogram incl. 5-part diff.
HOR12	Hormones
INF12	Inf. Disease Combination Control
SP12	Specific Proteins
SYP12	Syphilis Serology
TM12	Tumor Marker
TMH12	Tumor Marker & Hormones

Bimonthly	Program / Date*
	25/02/2025 - 11/03/2025
	22/04/2025 - 06/05/2025
	17/06/2025 - 01/07/2025
	19/08/2025 - 02/09/2025
	21/10/2025 - 04/11/2025
	02/12/2025 - 16/12/2025
CASE-T	Case Studies in Clin. Laboratory Science

Quarterly	Program / Date*
	18/02/2025 - 11/03/2025
	15/04/2025 - 06/05/2025
	15/07/2025 - 05/08/2025
	14/10/2025 - 04/11/2025
AB0	Blood grouping
ANTI-THYR	Thyroid antibodies
BAC	Bacteriology
BG4	Blood Gas and Electrolytes
BILI-N	Bilirubin neonatal
CC4	Clinical Chemistry
CM4	Cardiac Marker
COA4	Coagulation
COVAG	SARS-CoV-2 Antigen
COVM	SARS-CoV-2 Molekular
CSF4	CSF diagnostics
DAT	Drugs of Abuse
EBV	Epstein-Barr Virus Serology
ESR	Erythrocyte sedimentation rate
ETH4	Ethanol, Ammonia and Bicarbonate
GHB4	Glycated Hemoglobin (HbA1c)
GLUWB	Glucose POC - Whole Blood
HAV	Hepatitis A Virus Serology
HBV	Hepatitis B Virus Serology
HBVM	HBV Molecular
HCG	hCG in serum
HCGU	hCG in urine
HCVM	HCV Molecular
HEM3D	Hemogram incl. 3-part diff.
HEM4	Hemogram
HEM5D4	Hemogram incl. 5-part diff.
HIV	HIV Antibodies and Antigen
HIVM	HIV Molecular
HOR4	Hormones
INF4	Inf. Disease Combination Control
INF4x4	Inf. Disease Combination Control
INR-POCT	Prothrombin Time (POCT)
MALM	Malaria Microscopy
OXI4	Co-Oximetry
PCT	Procalcitonin
SP4	Specific Proteins
SYP4	Syphilis Serology
TDM	Therapeutic Drug Monitoring
TM4	Tumor Marker
TMH4	Tumor Marker & Hormones
TORCH	ToRCH Serology
UC	Urine Chemistry
US4	Qualitative Urine Analysis (Urine stick)
USEDL4	Urine Sediment for light scattering methods
USEDM4	Urine Sediment for microscopic methods

* Start of the measurement period until closing date

Registration deadline: in each case 3 months before the start of the corresponding measurement period.

Late registrations can still be considered if samples are available.

Semi-annual 1 (Q1+Q3)	Program / Date*	Semi-annual 2 (Q2+Q4)	Program / Date*
	18/02/2025 - 11/03/2025		29/04/2025 - 20/05/2025
	15/07/2025 - 05/08/2025		28/10/2025 - 18/11/2025
CC2	Clinical Chemistry	ADE	Adenovirus Serology
CM2	Cardiac Marker	ASF	Aspergillus Fumigatus Serology
COA2	Coagulation	ASPAG	Aspergillus Galactomannan Antigen
HEM2	Hemogram	BOR	Borrelia Serology
IMHEM	Immunohematology	BOR-G-AI	Borrelia IgG antibody index
INF2	Inf. Disease Combination Control	BOR-M-AI	Borrelia IgM antibody index
SYP2	Syphilis Serology	BPES	Bordetella Serology
TMH2	Tumor Marker & Hormones	BRU	Brucella Serology
US2	Qualitative Urine Analysis (Urine stick)	CHA	Chagas Serology
USEDL2	Urine Sediment for light scattering methods	CHIKV	Chikungunya Virus Serology
USEDM2	Urine Sediment for microscopic methods	CHT	Chlamydia Trachomatis Serology
		COVID	SARS-CoV-2 Serology
		COX	Coxsackievirus Serology
		DENV	Dengue Virus Antibodies
		DENVAG	Dengue Virus NS1 Antigen
		ECH	ECHO Virus Serology
		ENT	Enterovirus Serology
		ESRAF	Erythrocyte sedimentation rate for Alifax
		ESRAL	Erythrocyte sedimentation rate for Alcor iSED analysers
		FLUAAG	Influenza A Antigen Detection
		FLUBAG	Influenza B Antigen Detection
		FOB	Fecal Occult Blood
		HEV	Hepatitis E Virus Serology
		HPYL	Helicobacter Pylori Antibodies
		HTL	HTLV I/II
		INA	Influenza A Virus Serology
		INB	Influenza B Virus Serology
		LEP	Leptospira Serology
		LPAB	Legionella Pneumophila Antibodies
		MEA	Measles Serology
		MYPL	Mycoplasma Antibodies
		PAR	Parvovirus B19 Serology
		PIN	Parainfluenza Virus Serology
		RESPAG	Respiratory Viral Antigen Detection
		RSV	RSV Serology
		RSVAG	RSV Antigen Detection
		STAA	Streptococcus A Antigen
		TBEV-G-AI	TBEV IgG antibody index
		TBEV-M-AI	TBEV IgM antibody index
		VZV	Varizella Zoster Virus Serology
		WNV	West Nile Virus Serology
		ZIKV	Zika Virus Serology

* Start of the measurement period until closing date

Registration deadline: in each case 3 months before the start of the corresponding measurement period.

GENERAL TERMS FOR THE PARTICIPATION IN EXTERNAL QUALITY ASSESSMENT SURVEYS OF ESFEQA

Status July 2024

1. Participation
Participation in ESfEQA external quality assessment (EQA) surveys is open to anyone who performs laboratory tests in their own practice or in a managed medical laboratory. The following conditions for participation apply.

2. Consent to conditions of participation
By registering with ESfEQA GmbH, the participant agrees to these general terms and conditions of participation.

3. Assignment of services
Individual elements of EQA schemes (e.g. pretesting of values, packaging and shipping) may be assigned to subcontractors. ESfEQA is responsible for the work of the subcontractors.

4. ESfEQA catalog
The ESfEQA portfolio of EQA schemes and analytes contained in individual programs are described in the ESfEQA catalog. Depending on the availability of samples and number of participants, ESfEQA reserves the right not to offer the entire spectrum of analytes for each EQA survey or sample.

5. Schedule
The schedule, published in the ESfEQA catalog, contains deadlines for ordering and result submission, as well as the testing periods. Once the deadline for ordering has passed, acceptance of late orders is at ESfEQA's discretion. Results must be submitted to ESfEQA electronically, or using a result entry form, on or before the closing date. All deadlines and calendar dates are in the same time zone as ESfEQA's place of business in Heidelberg, Germany (i.e. GMT+1).

6. Cancelation of EQA surveys
ESfEQA reserves the right to cancel or postpone EQA surveys. This information will be provided to participants before the original sample shipping date and ESfEQA will endeavor to offer an alternative date in a timely manner.

7. Registration
For participation in ESfEQA EQA surveys registration is required. This can be done online, or by sending the necessary information to ESfEQA by email to: surveys@esfeqa.eu. The following information is required: laboratory name, name of the organization/hospital, name of participant, number of analytical devices, and e-mail address.

8. Ordering of samples
Distribution of ESfEQA EQA surveys is usually carried out by international distributors. If there is no distributor available in the participant's region, sales can be carried out directly by ESfEQA. The ordering process between participants and distributors is the responsibility of the parties involved. As a rule, an EQA program is ordered for a full calendar year. Orders placed during the year generally include the survey samples up to the end of the current calendar year.

9. Homogeneity and stability of EQA samples
The EQA survey samples selected by ESfEQA were examined and evaluated with regard to homogeneity and stability.
report display, the upper limit of the permissible range corresponds to a z-value of 3 and the lower limit to a z-value of -3.

10. Designation of EQA samples
The EQA samples can be distinguished by their identifier, which has the following format: program acronym_survey year_survey number_sample number. For example, the sample CM4_2025_01_a belongs to the quarterly program Cardiac Marker (CM4) in the year 2025 and is sample "a" of the first survey. Samples with the same designation are not necessarily identical, i.e. different results can be obtained despite the same designation. ESfEQA correctly allocates samples to the original batch and thus to the target values.

11. Shipping of EQA samples
EQA samples are shipped by postal or parcel service. Due to governmental restrictions, or insufficient sample stability, shipping of individual EQA programs to specific countries may be excluded.

12. Instructions for Use
Instructions for Use (IFU) for each EQA survey are available on the ESfEQA website (www.esfeqa.eu). A printout of the IFU is usually enclosed with the sample package. Each IFU includes instructions for sample preparation and stability.

13. Use of EQA samples
Usually, EQA samples should be treated exactly like patient samples, measured in the same way as routine samples according to manufacturer's instructions for instruments and reagents. They may only be used for the purpose of participating in an EQA survey and may not be used in a misappropriated manner. Generally normal laboratory procedure for testing potentially hazardous and potentially infectious samples also applies to EQA samples.

14. Submission of survey results
Where applicable, submission of results includes the actual measured value as well as method, instrument and reagent used. The input mask in TEQA (ESfEQA's evaluation software application) displays the required information for each EQA program. A drop-down list of methods, instruments and reagents is provided in the configuration section.
If a participant's method, instrument or reagent is not listed in TEQA, participants can add this information using the in¬put mask "coding request". They can select their specific method, instrument and reagent to create a new configuration prior to entering their test results.
The selection of method, instrument and reagent, as well as submission of results, must be performed using the TEQA web-application. Participants receive their login data (username and password) from ESfEQA, which is required to enter results. The password consists of at least 8 characters, including at least 2 special characters. Username and password are to be treated confidentially by the participant.
An alternative to result submission via the TEQA web-application, is the result form, that can be sent to ESfEQA either by E-Mail (info@esfeqa.eu) or Fax (+49 6221 894669-90). Result forms specific for each EQA program are provided on the ESfEQA website. ESfEQA encourages all participants to submit their results online via the secured TEQA web-application, for the sake of data security and convenience.
ESfEQA evaluates all survey results submitted by participants by the deadline. In the event of loss or late arrival of their data the participant bears the risk. ESfEQA are not obliged to evaluate results submitted after the

submission deadline.
Quantitative results are generally reported with a value and a unit. The participant determines the number of digits for reporting. In general, results should be reported as measured. However, results submitted as “< test range’ (e.g. “< 10”) or “> test range” (e.g. “>2000“) are not valid.
For results below the test range, the lower test range limit should be reported (e.g. “10”).
For samples with analyte concentrations above the test range, the sample can be diluted (if recommended for particular applications) or the upper test range limit (e.g. “2000”) can be submitted as the result. Several units are usually available for entering quantitative results. The units are converted into the standard unit used by ESfEQA.
Laboratories are obliged to treat their results confidentially and not to pass them on to third parties until the EQA survey report has been received. If ESfEQA becomes aware of the passing on or falsification of results or the collusion between participants, ESfEQA reserves the right to exclude those concerned from further participation in EQA surveys conducted by ESfEQA as well as to exclude the issuance of reports.

15. Number of results per participant
For each EQA sample and analyte, up to 3 values per participant can be submitted. The values have to be determined by different analytical devices that are independent from each other.

16. Correction of transmitted results
Participants can edit their results, using a change request via the TEQA web-application, up until the deadline for result submission of the EQA survey. ESfEQA must check and accept change requests before results are edited accordingly. A change re¬quest can also be submitted by participants via e-mail or fax to ESfEQA, up until the deadline for result submission. Participants who submitted their results via the TEQA web-application can only use a change request via the TEQA web application.

17. Evaluation of EQA results
For each analyte in ESfEQA EQA surveys, the type of target value determination and acceptance criterion are predefined. For quantitative parameters, the target value is usually the consensus value of participants results. This value is calculated according to ISO 13528:2022-08 'Statistical methods for use in proficiency testing by interlaboratory comparison' using robust statistics.
Samples provided for testing of qualitative parameters are thoroughly tested with different analytical systems before being used as control material, thereby setting the target value.
System-specific differences are taken into account, if appropriate and feasible, with a corresponding statistical evaluation. The broadest possible distinction according to method, device and/or reagent used, is made available to participants (M-, I-, R-group). The minimum number of results of an evaluation group is 5. If this number is not reached in the survey, the individual result has to be compared to the robust mean of the next largest group that can be evaluated. Usually, this is the group consisting of participants using the same method (M group), or the general group containing the results of all participants. The definition of the evaluation group is documented in the survey report.
For quantitatively determined analytes, the maximum permissible ranges of the target value are predefined. The permissible range for each analyte is derived from its medical relevance as well as the reference interval. In the report display, the upper limit of the permissible range corresponds to a z-value of 3 and the lower limit to a z-value of -3.

18. Survey reports
In general, the participants will receive reports electronically via the TEQA web-application within 10 days (for monthly programs), or three weeks (for quarterly and semi-annual programs) after the deadline for submission of the results. The reports include the results submitted by the participant evaluated in comparison to target values. The data is displayed both in tabular and illustrated form (e. g., Histogram, Shewart chart, Youden plot). The reports are intended for external quality assurance of laboratories. They may not be published, passed on or used for purposes other than quality assurance without the written consent of ESfEQA.

19. Fees
The fees for the participation are set and communicated to the participants by the responsible distributor of ESfEQA programs in their geographical area/country. Due to the high variability of shipping, handling and customs costs, the prices may vary between various countries. Please contact your distributor for the participation fee.

20. Certificates
For each EQA program participants receive a certificate of participation. In addition, participants receive a certificate for those parameters which met the specified performance criteria in the respective EQA survey. Both certificates are made available to participants via the TEQA web-application. The certificates are issued simultaneously with the reports.

21. Loss and damage of EQA test material
In the event of sample loss or damage, ESfEQA should be notified immediately. If possible, ESfEQA will send replacement samples without acknowledging any claims. However, the contract is fulfilled on the date of dispatch of the original sample material.

22. Complaints and appeals
After receipt of an EQA survey report, a complaint/appeal can be made within a period of 4 weeks. After expiry of this period, any claims by the participant based on a complaint /appeal are excluded. In the event of a justified complaint/ appeal, ESfEQA will decide whether to reimburse the amount paid for the EQA survey, or to provide a substitute EQA survey. ESfEQA GmbH does not reimburse any costs incurred for reagents, time expenditure etc. unless ESfEQA GmbH is liable in accordance with paragraph 23 of these General Terms and Conditions for Participation.

23. Warranty
ESfEQA shall only be liable for damages of any kind in the case of intent and gross negligence, if the other prerequisites for claims are met. In all other respects, liability for damages of any kind, regardless of the basis of the claim, including liability for culpa in contrahendo, is excluded.

24. Confidentiality
Individual EQA data is kept confidential. It is only known to the corresponding participant, their distributor and ESfEQA employees. ESfEQA collects, processes and uses personal data of the participant only to the extent necessary for the performance of EQA surveys, the preparation of the reports and for the purpose of quality assurance. This includes the forwarding of the data identifiable by subscriber and device number for quality assurance measures to the respective manufacturer of the analytical systems (device and reagent).

COMPANY INFORMATION

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